

OVERVIEW

Reports from the fifth edition of CAGI: The Critical Assessment of Genome Interpretation

Gaia Andreoletti¹  | Lipika R. Pal²  | John Moulton^{2,3}  | Steven E. Brenner^{1,4} 

¹Department of Plant and Microbial Biology, University of California, Berkeley, California

²Institute for Bioscience and Biotechnology Research, University of Maryland, Rockville, Maryland

³Department of Cell Biology and Molecular Genetics, University of Maryland, College Park, Maryland

⁴Center for Computational Biology, University of California, Berkeley, California

Correspondence

John Moulton, Institute for Bioscience and Biotechnology Research, University of Maryland, 9600 Gudelsky Drive, Rockville, MD 20850.

Email: jmoulton@umd.edu

Steven E. Brenner, Department of Plant and Microbial Biology, University of California, Berkeley, CA 94720.

Email: brenner@compbio.berkeley.edu

Funding information

National Human Genome Research Institute and National Cancer Institute, Grant/Award Number: U41 HG007346 and R13 HG006650

Abstract

Interpretation of genomic variation plays an essential role in the analysis of cancer and monogenic disease, and increasingly also in complex trait disease, with applications ranging from basic research to clinical decisions. Many computational impact prediction methods have been developed, yet the field lacks a clear consensus on their appropriate use and interpretation. The Critical Assessment of Genome Interpretation (CAGI, /'kɑ-jē/) is a community experiment to objectively assess computational methods for predicting the phenotypic impacts of genomic variation. CAGI participants are provided genetic variants and make blind predictions of resulting phenotype. Independent assessors evaluate the predictions by comparing with experimental and clinical data.

CAGI has completed five editions with the goals of establishing the state of art in genome interpretation and of encouraging new methodological developments. This special issue (<https://onlinelibrary.wiley.com/toc/10981004/2019/40/9>) comprises reports from CAGI, focusing on the fifth edition that culminated in a conference that took place 5 to 7 July 2018. CAGI5 was comprised of 14 challenges and engaged hundreds of participants from a dozen countries. This edition had a notable increase in splicing and expression regulatory variant challenges, while also continuing challenges on clinical genomics, as well as complex disease datasets and missense variants in diseases ranging from cancer to Pompe disease to schizophrenia. Full information about CAGI is at <https://genomeinterpretation.org>.

KEYWORDS

CAGI, Critical Assessment of Genome Interpretation, genomics, genetic variation, cancer genetics, variant impact predictors, SNP

1 | INTRODUCTION

Interpretation of genome sequence variation plays a major and increasing role in both basic research and in clinical medicine. These applications necessitate robust and reliable computational approaches to aid in determining the phenotypic impact of variants. Many such methods have been developed (see Hu et al., 2019 in this issue). However, the appropriate use and accuracy of most methods have not been objectively determined.

The Critical Assessment of Genome Interpretation (CAGI, pronounced /'kɑ-jē/) addresses this need by providing an objective evaluation of the state-of-the-art in relating human genetic variation to phenotype, particularly health. Each edition of CAGI provides about a dozen challenges to understand performance of prediction methods in a given scenario. Participants in a challenge are provided genetic variants and make predictions of resulting molecular, cellular, or organismal phenotypes. Data sets for the challenges include germline and somatic cancer variation, rare disease, common disease, and pharmacogenomics. The scale of challenges ranges from single

nucleotides to whole genomes, as well as complementary multiomics and environmental information. Variant types include those affecting expression, splicing, and amino acid sequence, and may be single base changes, insertions or deletions, and structural variation.

The CAGI cycle commences with the organizers soliciting data sets from researchers and clinicians, whose studies demonstrate relationships between genotype and phenotype. Such data sets must be fully developed (“ripe”) but not publicly available (“spoiled”) until after the CAGI prediction season. Data providers and organizers work together to develop these data sets into prediction challenges with well-defined data sets and goals. The CAGI Ethics Forum evaluates and adjusts these challenges for suitability and sensitivity. In parallel, predictors are enrolled, bound by the CAGI Data Use Agreement [<https://genomeinterpretation.org/data-use-agreement>], and vetted for access to the CAGI experiment with tiers reflecting the sensitivity of the data. The prediction season launches with release of the challenges and concludes with the submission of predictions from predictors. For the fifth edition of CAGI (CAGI5), the prediction season extended until May 2018. The submitted predictions are evaluated by independent assessors. Each CAGI edition culminates in a conference to discuss the outcome; the CAGI5 conference was held from 5 to 7 July 2018, for which videos and slide sets are publicly available to registered CAGI members. [<https://genomeinterpretation.org/content/5-conference>]

Results from the previous CAGI edition (CAGI4) were published in a special issue of this journal (Hoskins et al., 2017). The present issue contains assessment and participant papers primarily from the most recent experiment, CAGI5, which included 14 challenges and attracted participants from 12 countries.

Notable in this CAGI edition is the increasing representation of regulatory and noncoding variant challenges. Previously, CAGI has had just one small splicing challenge [<https://genomeinterpretation.org/content/Splicing-2012>]. CAGI5 included two full-scale splicing challenges (Mount et al., 2019) and these have resulted in five papers from participants (Chen, Lu, Zhao, & Yang, 2019; Cheng, Çelik, Nguyen, Avsec, & Gagneur, 2019; Gotea, Margolin, & Elnitski, 2019; Naito, 2019; Wang, Wang, & Hu, 2019). The issue also contains an overview paper from one of the splicing data providers (Rhine et al., 2019). There had also been only one previous expression regulatory variant challenge, in CAGI4 (Kreimer et al., 2017). CAGI5 has a new expression regulatory challenge (Shigaki et al., 2019), and there are also two participant papers (Dong & Boyle, 2019; Kreimer, Yan, Ahituv, & Yosef, 2019).

CAGI5 continued the emphasis on the interpretation of clinically relevant large-scale sequence data, with a challenge on the risk of thrombosis in African-American cohort given whole exome sequence (McInnes et al., 2019; Wang & Bromberg, 2019); the identification of variants contributing to intellectual disability phenotypes given gene panel sequence (Aspromonte et al., 2019; Carraro et al., 2019; Chen, 2019); and a challenge of matching whole genome sequences to clinical profiles for patients at Toronto’s Hospital for Sick Children (SickKids) and identifying causal variants (Kasak, Hunter et al., 2019; Pal, Kundu, Yin, & Moul, 2019). The latter challenge is related to the

CAGI4 SickKids challenge, also described in the assessment paper here.

Over all CAGI editions, the plurality of challenges have been on the interpretation of isolated missense variants, and CAGI5 continues that trend. There are assessment, data provider, and participant papers for the prediction of the destabilizing effect of missense mutations in a cancer-relevant protein (Frataxin, with biophysical measurements of protein stability; Petrosino et al., 2019; Savojardo, Petrosino et al., 2019; Strokach, Corbi-Verge, & Kim, 2019); on the effect of missense changes in a human calmodulin, assayed using a high-throughput yeast complementation assay (Zhang et al., 2019); the effect of missense mutations related to schizophrenia in human Pericentriolar Material 1 (*PCM1*), using a zebrafish development model (Miller, Wang, & Bromberg, 2019; Monzon et al., 2019); the effect of missense mutations in two cancer-related proteins, *PTEN* and *TPMT*, on intracellular protein levels, measured in a high-throughput assay (Pejaver et al., 2019); and the effect of missense changes in a monogenic disease related protein, acid alpha-glucosidase (*GAA*), with measurements of total intracellular enzyme activity (Adhikari, 2019). Three participant papers describe results on all the missense challenges (Garg & Pal, 2019; Katsonis & Lichtarge, 2019; Savojardo, Babbi et al., 2019). The issue also contains assessment articles from two earlier missense challenges on monogenic disease related proteins: N-acetyl-glucosaminidase (*NAGLU*; Clark et al., 2019), with total intracellular enzyme activity measured; and cystathionine beta-synthase (*CBS*), using the metric of yeast growth in a complication assay (Kasak, Bakolitsa et al., 2019).

In addition to the other cancer-related challenges outlined above, there are two that required prediction of the pathogenicity of germline variants in cancer-related proteins: one for breast cancer risk from variants in *BRCA1* and *BRCA2* as characterized by the ENIGMA consortium (Cao et al., 2019; Cline et al., 2019; Padilla et al., 2019; Parsons et al., 2019), and the other for cancer risk of variants in *CHEK2* in Latina breast cancer cases and ancestry matched controls (Voskanian et al., 2019).

CAGI5 will continue to bear fruit. This edition introduced a time-based challenge in association with dbNSFP, whereby CAGI accepted predictions for all possible missense variants in the genome. The results are to be vetted periodically in the future as the impact of some of these variants are experimentally or clinically established. Additional papers on the challenges and other aspects of CAGI are presently in development and will be added to the CAGI5 papers collection at *Human Mutation* (<https://onlinelibrary.wiley.com/toc/10981004/2019/40/9>).

Full information about CAGI5 and earlier editions is at <https://genomeinterpretation.org>.

ACKNOWLEDGMENTS

The primary contributors to CAGI are predictors, and without their willingness to test their methods in this way, the experiments would not be possible. Participants over the five CAGI editions are: Allison

Abad, Ogun Adebali, Aashish Adhikari, Ivan Adzhubey, Talal Amin, Žiga Avsec, Johnathan R. Azaria, Giulia Babbi, Eraan Bachar, Benjamin Bachman, Minkyung Baek, Greet De Baets, Michael A. Beer, Violeta Beleva-Guthrie, Riccardo Bellazzi, Bonnie Berger, Brady Bernard, Rajendra R. Bhat, Rohit Bhattacharya, Samuele Bovo, Alan P. Boyle, Marcus Breese, Aharon S. Brodie, Yana Bromberg, Binghuang Cai, Colin Campbell, Chen Cao, Yue Cao, Emidio Capriotti, Liran Carmel, Marco Carraro, Hannah Carter, Rita Casadio, Muhammed Hasan Çelik, Billy H. W. Chang, Chien-Yuan Chen, Haoran Chen, Jingqi Chen, Ken Chen, Shann-Ching Chen, Yun-Ching Chen, Jun Cheng, Melissa Cline, Noa Cohen, Carles Corbi-Verge, Andrea Corredor, Chen Cui, Dvir Dahary, Carla Davis, Xavier de la Cruz, Mark Diekhans, Rezarta I. Dogan, Shengcheng Dong, Christopher Douville, Ian Driver, Roland Dunbrack, Joost van Durme, Orland Diez, Andrea Eakin, Matthew Edwards, Laura Elnitski, Gokcen Eraslan, Hai Fang, Alexander V. Favorov, Tzila Fenesh, Xin Feng, Carlo Ferrari, Simon Fishilevich, Anna Flynn, Lukas Folkman, Colby T. Ford, Adam Frankish, Zaneta Franklin, Yao Fu, Julien Gagneur, Aditi Garg, Alessandra Gasparini, Tom Gaunt, David Gifford, Manuel Giollo, Nina Gonzaludo, Valer Gotea, Julian Gough, Solomon Grant, Rafael Guerrero, Yuchun Guo, Sara Gutiérrez-Enríquez, James Jisu Han, Jennifer Harrow, Marcia Hasenahuer, Lim Heo, Tamar Holzer, Ramin Homayouni, Alex Hawkins-Hooker, Raghavendra Hosur, Cheng L. V. Huang, Chad D. Huff, Peter Huwe, Sohyun Hwang, Tadashi Imanishi, Jules Jacobsen, Chan-Seok Jeong, Yuxiang Jiang, David T. Jones, Daniel Jordan, Thomas Joseph, Beomchang Kang, Rachel Karchin, Mostafa Karimi, Panagiotis Katsonis, Sunduz Keles, Manolis Kellis, Henry Kenlay, Nikki Kiga, Dongsup Kim, Eiru Kim, Philip M. Kim, Jack F. Kirsch, Michael Kleymann, Sujatha Kotte, Andreas Kraemer, Anat Kreimer, Ivan Kulakovskiy, Anshul Kundaje, Kunal Kundu, Pui-Yan Kwok, Carmen Lai, Ernest Lam, Doron Lancet, Dae Lee, Gyu Rie Lee, Insuk Lee, Pietro Di Lena, Emanuela Leonardi, Andy Li, Biao Li, Mulin Jun Li, Yue Li, Olivier Lichtarge, Ivan Limongelli, Chiao-Feng Lin, Qewang Liu, Rhonald C. Lua, Angel Mak, Vsevolod J. Makeev, Gennady Margolin, Pier Luigi Martelli, David Masica, Zev Medoff, Aziz M. Mezlini, Maximilian Miller, Gilad Mishne, Rahul Mohan, Alejandro Moles-Fernández, Gemma Montalban, Alexander M. Monzon, Sean D. Mooney, Oluwaseyi Adebayo Moronfoye, Matthew Mort, John Moulton, Steve Mount, Eliseos Mucaki, Jonathan Mudge, Nikola Mueller, Chris Mungall, Katsuhiko Murakami, Yoko Nagai, Tatsuhiko Naito, Thi Yen Duong Nguyen, Giovanna Nicora, Noushin Niknafs, Abhishek Niroula, Conor M. L. Nodzak, Robert O'Connor, Yanay Ofran, Ayodeji Olatubosun, Lars Ootes, Selen Özkan, Kivilcim Ozturk, Natàlia Padilla, Kymberleigh Pagel, Debnath Pal, Lipika R. Pal, Taeyong Park, Nathaniel Pearson, Vikas Pejaver, Jian Peng, Dmitry D. Penzar, Alexandra Piryatinska, Catherine Plotts, Jennifer Poitras, Predrag Radivojac, Sadhna Rana, Aditya Rao, Aliz R. Rao, Sandra Bonache Real, John Reid, Casandra Riera, Graham Ritchie, Ettore Rizzo, Peter Rogan, Frederic Rousseau, Vangala Govindakrishnan Saipradeep, Castrense Savojardo, Jana M. Schwarz, Joost Schymkowitz, Chaok Seok, George Shackelford, Sohela Shah, Maxim Shatsky, Yang Shen, Yuan Shi, Ron Shigeta, Hashem A. Shihab, Jung E. Shim, Junha Shin, Sunyoung Shin, Ilya

Shmulevich, Bradford R. Silver, Nasa Sinnott-Armstrong, Vasily V. Sitnik, Naveen Sivadasan, Ben Smithers, Yeşim A. Son, Rajgopal Srinivasan, Mario Stanke, Nathan Stitzel, Alexey Strokach, Andrew Su, Yuanfei Sun, Lakshman Sundaram, Uma Sunderam, Paul Tang, Sean V. Tavtigian, Nuttinee Teerakulkittipong, Natalie Thurlby, Janita Thusberg, Kevin Tian, Collin Tokheim, Scott Topper, Silvio C. E. Tosatto, Yemliha Tuncel, Tychele Turner, Ron Unger, Aneeta Uppal, Gurkan Ustunkar, Jouni Valiaho, Mauno Vihinen, Ilya E. Vorontsov, Mary Wahl, Michael Wainberg, Li-San Wang, Maggie Wang, Meng Wang, Robert Wang, Xinyuan Wang, Yanran Wang, Liping Wei, Qiong Wei, Rene Welch, Stephen Wilson, Chunlei Wu, Lijing Xu, Qifang Xu, Zhongxia Yan, Yang Yang, Yuedong Yang, Zhaomin Yao, Christopher Yates, Yizhou Yin, Nir Yosef, Erin Young, Chen-Hsin Yu, Yao Yu, Dejian Yuan, Jan Zaucha, Haoyang Zeng, Fengfeng Zhou, Yaoqi Zhou, and Maya Zuhl.

CAGI also depends critically on the generosity of the groups who make their data available and help develop the challenges, usually before their own publications: Nadav Ahituv, Russ Altman, Adam P. Arkin, Madeleine P. Ball, Jason Bobe, Paolo Bonvini, Bethany Buckley, Roberta Chiaraluce, George Church, Wyatt T. Clark, Valerio Consalvi, The ENIGMA Consortium, Garry R. Cutting, Emma D'Andrea, Roxana Daneshjou, Lisa Elefanti, Will Fairbrother, Aron W. Fenton, Douglas M. Fowler, Andre Franke, David Goldgar, Nina Gonzaludo, Brenton R. Graveley, Joe W. Gray, Linnea Jansson, John P. Kane, Nicholas Katsanis, Martin Kircher, Pui-Yan Kwok, Rick Lathrop, Jonathan H. LeBowitz, Emanuela Leonardi, Xiaoming Liu, Federica Lovisa, Angel C. Y. Mak, Mary J. Malloy, Kenneth Matreyek, Christian Marshall, Richard McCombie, Chiara Menin, M. Stephen Meyn, John Moulton, Alessandra Murgia, Thomas Nalpathamkalam, Robert L. Nussbaum, Lipika R. Pal, Michael Parsons, Britt-Sabina Petersen, Mehdi Pirooznia, James B. Potash, Clive R. Pullinger, Jasper Rine, Frederick P. Roth, Kevin Ru, Pardis Sabeti, Jeremy Sanford, Maria C. Scaini, Nicole Schmitt, Jay Shendure, Molly Sheridan, Michael Snyder, Amanda Spurdle, Lea Starita, Wilson Sung, Bhooma Thiruvahindrapuram, Tim Sterne-Weiler, Joe Whitney, Paul L. F. Tang, Sean Tavtigian, Ryan Tewhey, Silvio C. E. Tosatto, Jochen Weile, G. Karen Yu, Peter Zandi, and Elad Ziv.

We offer great thanks to the CAGI assessors who have devoted extensive time to analysis and evaluation of the results: Aashish Adhikari, Michael A. Beer, Yana Bromberg, Emidio Capriotti, Marco Carraro, John-Marc Chandonia, Rui Chen, Luigi Chiricosta, Wyatt T. Clark, Melissa Cline, Roxana Daneshjou, Roland Dunbrack, Iddo Friedberg, Mabel Furutsuki, Gad Getz, Manuel Giollo, Nick Grishin, Maricel Kann, Rachel Karchin, Anat Kreimer, Greg McInnes, M. Stephen Meyn, Sean D. Mooney, Alexander A. Morgan, John Moulton, Steve Mount, Robert L. Nussbaum, Ayoti Patra, Francesco Reggiani, Jeremy Sanford, David B. Searls, Dustin Shigaki, Artem Sokolov, Josh Stuart, Shamil Sunyaev, Sean Tavtigian, Silvio C. E. Tosatto, Alin Voskanyan, Qifang Xu, and Nir Yosef.

Our advisory board and scientific council members have been the source of much wisdom and assistance. The board has comprised: Russ Altman, Serafim Batzoglou, George Church, Tim Hubbard, Scott Kahn, Rachel Karchin, Sean D. Mooney, Pauline Ng, Robert L. Nussbaum, Susanna Repo, John Shon, and Michael Snyder; our Scientific Council

additionally has included: Patricia Babbitt, Yana Bromberg, Atul Butte, Garry R. Cutting, Laura Elnitski, Reece Hart, Ryan Hernandez, Michael Snyder, Shamil Sunyaev, Joris Veltman, Liping Wei, and Justin Zook. The ethicists and patient advocates of the CAGI Ethics Forum have worked with organizers, undertaking a critical role in ensuring CAGI operates on a firm ethical foundation and informing its activities: Wylie Burke, Larry Carr, Flavia Chen, Steve Grossman, Julie Harris-Wai, Kirsten Isgro, Barbara A. Koenig, Selena Martinez, Robert L. Nussbaum, Jodi Paik, and Mark Yarborough.

We also thank those who have helped organize the CAGI experiments and who assisted with the associated technology: Daniel Barsky, John-Marc Chandonia, Ajithavalli Chellappan, Flavia Chen, Navya Dabhiru, Reece Hart (who coined the term “CAGI”), Roger Hoskins, Naveen Kollipara, Melissa K. Ly, Andrew J. Neumann, Gaurav Pandey, Sadhna Rana, Susanna Repo, Rajgopal Srinivasan, Stephen Yee, Sri Jyothsna Yeleswarapu, and Maya Zuhl. We specially thank those who helped in writing several papers for CAGI5 special issue: Constantina Bakolitsa, Zhiqiang Hu, and Laura Kasak.

We are truly grateful to NHGRI and NCI for the support of the CAGI experiment and conference. We especially recognize Tata Consultancy Services (TCS), which has been a collaborator in organizing all the CAGI experiments. We also thank the members of the Brenner and Moulton laboratories who have contributed to CAGI.

For this special issue we greatly appreciate the efforts of the anonymous peer reviewers, the Lead Guest Editor Rachel Karchin, and the Consulting Guest Editors who oversaw the peer-review process: Tim Hubbard, Sean D. Mooney, Robert L. Nussbaum, Predrag Radivojac, Shamil Sunyaev, and Joris Veltman. John Moulton and Steven E. Brenner were the Issue Editors, and Lipika R. Pal was the Organizing Editor for this special issue of *Human Mutation*. We thank Mohammad Israr, Christine Murray, Mark Paalman, and Allain Selga for their work in coordinating the editorial and production operations at Wiley.

Finally, we also wish to acknowledge our great debt to the many individuals who shared their private genetic and phenotypic or clinical information as participants in the research studies and clinical data sets included in the CAGI challenges.

CONFLICT OF INTERESTS

There is no conflict of interest involved in this study.

FUNDING

The CAGI experiment coordination is supported by NIH / NHGRI and NCI U41 HG007346 and the CAGI conference by R13 HG006650. Support has also been provided via a research agreement with Tata Consultancy Services (TCS).

ORCID

Gaia Andreoletti  <http://orcid.org/0000-0002-0452-0009>

Lipika R. Pal  <http://orcid.org/0000-0002-3390-110X>

John Moulton  <http://orcid.org/0000-0002-3012-2282>

Steven E. Brenner  <http://orcid.org/0000-0001-7559-6185>

REFERENCES

- Adhikari, A. N. (2019). Gene-specific features enhance interpretation of mutational impact on acid alpha-glucosidase enzyme activity. *Human Mutation*, <https://doi.org/10.1002/humu.23846>.
- Aspromonte, M. C., Bellini, M., Gasparini, A., Carraro, M., Bettella, E., Polli, R., ... Leonardi, E. (2019). Characterization of Intellectual disability and Autism comorbidity through gene panel sequencing. *Human Mutation*, <https://doi.org/10.1002/humu.23822>
- Cao, Y., Sun, Y., Karimi, M., Chen, H., Moronfoye, O., & Shen, Y. (2019). Predicting pathogenicity of missense variants with weakly supervised regression. *Human Mutation*, <https://doi.org/10.1002/humu.23826>
- Carraro, M., Monzon, A. M., Chiricosta, L., Reggiani, F., Aspromonte, M. C., Bellini, M., ... Leonardi, E. (2019). Assessment of patient clinical descriptions and pathogenic variants from gene panel sequences in the CAGI-5 intellectual disability challenge. *Human Mutation*, <https://doi.org/10.1002/humu.23823>
- Chen, J. (2019). A fully-automated event-based variant prioritizing solution to the CAGI5 intellectual disability gene panel challenge. *Human Mutation*, <https://doi.org/10.1002/humu.23781>
- Chen, K., Lu, Y., Zhao, H., & Yang, Y. (2019). Predicting the change of exon splicing caused by genetic variant using support vector regression. *Human Mutation*, <https://doi.org/10.1002/humu.23785>
- Cheng, J., Çelik, M. H., Nguyen, Y. D., Avsec, Ž., & Gagneur, J. (2019). CAGI5 splicing challenge: Improved exon skipping and intron retention predictions with MMSplice. *Human Mutation*, <https://doi.org/10.1002/humu.23788>
- Clark, W. T., Andreoletti, G., Kasak, L., Bakolitsa, C., Hu, Z., Babbi, G., ... LeBowitz, J. H. (2019). Assessment of predicted enzymatic activity of alpha-N-acetylglucosaminidase (NAGLU) variants of unknown significance for CAGI 2016. *Human Mutation*, <https://doi.org/10.1002/humu.23875>
- Cline, M. S., Babbi, G., Bonache, S., Cao, Y., Casadio, R., ... de la Cruz, X. ENIGMA Consortium (2019). Assessment of blind predictions of the clinical significance of BRCA1 and BRCA2 variants. *Human Mutation*, <https://doi.org/10.1002/humu.23861>
- Dong, S., & Boyle, A. P. (2019). Predicting functional variants in enhancer and promoter elements using RegulomeDB. *Human Mutation*, <https://doi.org/10.1002/humu.23791>
- Garg, A., & Pal, D. (2019). Exploring the use of molecular dynamics in assessing protein variants for phenotypic alterations. *Human Mutation*, <https://doi.org/10.1002/humu.23800>
- Gotea, V., Margolin, G., & Elnitski, L. (2019). CAGI experiments: Modeling sequence variant impact on gene splicing using predictions from computational tools. *Human Mutation*, <https://doi.org/10.1002/humu.23782>
- Hoskins, R. A., Repo, S., Barsky, D., Andreoletti, G., Moulton, J., & Brenner, S. E. (2017). Reports from CAGI: The Critical Assessment of Genome Interpretation. *Human Mutation*, *38*, 1039–1041. <https://doi.org/10.1002/humu.23290>
- Hu, Z., Yu, C., Furutsuki, M., Andreoletti, G., Ly, M., Hoskins, R., ... Brenner, S. E. (2019). VIPdb, a genetic variant impact predictor database. *Human Mutation*, <https://doi.org/10.1002/humu.23858>
- Kasak, L., Bakolitsa, C., Hu, Z., Rine, J., Dimster-Denk, D. F., Pandey, G., ... Friedberg, I. (2019). Assessing computational predictions of the phenotypic effect of cystathionine-beta-synthase variants. *Human Mutation*, <https://doi.org/10.1002/humu.23868>
- Kasak, L., Hunter, J. M., Udani, R., Bakolitsa, C., Hu, Z., Adhikari, A. N., ... Meyn, M. S. (2019). CAGI SickKids challenges: Assessment of phenotype and variant predictions derived from clinical and genomic data of children with undiagnosed diseases. *Human Mutation*, <https://doi.org/10.1002/humu.23874>

- Katsonis, P., & Lichtarge, O. (2019). CAGI5: Objective performance assessments of predictions based on the evolutionary action equation. *Human Mutation*, <https://doi.org/10.1002/humu.23873>
- Kreimer, A., Yan, Z., Ahituv, N., & Yosef, N. (2019). Meta-analysis of massively parallel reporter assays enables prediction of regulatory activity. *Human Mutation*, <https://doi.org/10.1002/humu.23820>
- Kreimer, A., Zeng, H., Edwards, M. D., Guo, Y., Tian, K., Shin, S., ... Yosef, N. (2017). Predicting gene expression in massively parallel reporter assays: A comparative study. *Human Mutation*, *38*, 1240–1250. <https://doi.org/10.1002/humu.23197>
- McInnes, G., Daneshjou, R., Katsonis, P., Lichtarge, O., Srinivasan, R. G., Rana, S., ... Altman, R. (2019). Predicting venous thromboembolism risk from exomes in the Critical Assessment of Genome Interpretation (CAGI) challenges. *Human Mutation*, <https://doi.org/10.1002/humu.23825>
- Miller, M., Wang, Y., & Bromberg, Y. (2019). What went wrong with variant effect predictor performance for the PCM1 challenge. *Human Mutation*, <https://doi.org/10.1002/humu.23832>
- Monzon, A. M., Carraro, M., Chiricosta, L., Reggiani, F., Han, J., Ozturk, K., ... Tosatto, S. C. E. (2019). Performance of computational methods for the evaluation of Pericentriolar Material 1 missense variants in CAGI-5. *Human Mutation*, <https://doi.org/10.1002/humu.23856>
- Mount, S. M., Avsec, Z., Carmel, L., Casadio, R., Celik, M. H., Chen, K., ... Zhao, H. (2019). Assessing predictions of the impact of variants on splicing in CAGI5. *Human Mutation*, <https://doi.org/10.1002/humu.23869>
- Naito, T. (2019). Predicting the impact of single nucleotide variants on splicing via sequence-based deep neural networks and genomic features. *Human Mutation*, <https://doi.org/10.1002/humu.23794>
- Padilla, N., Moles-Fernández, A., Riera, C., Montalban, G., Özkan, S., Ootes, L., ... de la Cruz, X. (2019). BRCA1- and BRCA2-specific in silico tools for variant interpretation in the CAGI 5 ENIGMA challenge. *Human Mutation*, <https://doi.org/10.1002/humu.23802>
- Pal, L. R., Kundu, K., Yin, Y., & Moul, J. (2019). Matching whole genomes to rare genetic disorders: Identification of potential causative variants using phenotype-weighted knowledge in the CAGI SickKids5 clinical genomes challenge. *BioRxiv*, 707687. <https://doi.org/10.1101/707687>
- Parsons, M. T., Tudini, E., Li, H., Hahnen, E., Wappenschmidt, B., Feliubadaló, L., ... Spurdle, A. B. (2019). Large scale multifactorial likelihood quantitative analysis of BRCA1 and BRCA2 variants: An ENIGMA resource to support clinical variant classification. *Human Mutation*, <https://doi.org/10.1002/humu.23818>
- Pejaver, V., Babbi, G., Casadio, R., Folkman, L., Katsonis, P., Kundu, K., ... Bromberg, Y. (2019). Assessment of methods for predicting the effects of PTEN and TPMT protein variants. *Human Mutation*, <https://doi.org/10.1002/humu.23838>
- Petrosino, M., Pasquo, A., Novak, L., Toto, A., Gianni, S., Mantuano, E., ... Consalvi, V. (2019). Characterization of human frataxin missense variants in cancer tissues. *Human Mutation*, <https://doi.org/10.1002/humu.23789>
- Rhine, C. L., Neil, C., Glidden, D. T., Cygan, K. J., Fredericks, A. M., Wang, J., ... Fairbrother, W. G. (2019). Future directions for high-throughput splicing assays in precision medicine. *Human Mutation*, <https://doi.org/10.1002/humu.23866>
- Savojardo, C., Petrosino, M., Babbi, G., Bovo, S., Corbi-verge, C., Casadio, R., ... Capriotti, E. (2019). Evaluating the predictions of the protein stability change upon single amino acid substitutions for the FXN CAGI5 challenge. *Human Mutation*, <https://doi.org/10.1002/humu.23843>
- Savojardo, C., Babbi, G., Bovo, S., Capriotti, E., Martelli, P. L., & Casadio, R. (2019). Are machine learning based methods suited to address complex biological problems? Lessons from CAGI-5 challenges. *Human Mutation*, <https://doi.org/10.1002/humu.23784>
- Shigaki, D., Adato, O., Adhikar, A. N., Dong, S., Hawkins-Hooker, A., Inoue, F., ... Beer, M. A. (2019). Integration of multiple epigenomic marks improves prediction of variant impact in saturation mutagenesis reporter assay. *Human Mutation*, <https://doi.org/10.1002/humu.23797>
- Strokach, A., Corbi-Verge, C., & Kim, P. M. (2019). Predicting changes in protein stability caused by mutation using sequence- and structure-based methods in a CAGI5 blind challenge. *Human Mutation*, <https://doi.org/10.1002/humu.23852>
- Voskanian, A., Katsonis, P., Lichtarge, O., Pejaver, V., Radivojac, P., Mooney, S. D., ... Kann, M. G. (2019). Assessing the performance of in-silico methods for predicting the pathogenicity of variants in the gene CHEK2, among Hispanic females with breast cancer. *Human Mutation*, <https://doi.org/10.1002/humu.23849>
- Wang, R., Wang, Y., & Hu, Z. (2019). Using secondary structure to predict the effects of genetic variants on alternative splicing. *Human Mutation*, <https://doi.org/10.1002/humu.23790>
- Wang, Y., & Bromberg, Y. (2019). Identifying mutation-driven changes in gene functionality that lead to venous thromboembolism. *Human Mutation*, <https://doi.org/10.1002/humu.23824>
- Zhang, J., Kinch, L. N., Cong, Q., Katsonis, P., Lichtarge, O., Savojardo, C., ... Grishin, N. V. (2019). Assessing predictions on fitness effects of missense variants in calmodulin. *Human Mutation*, <https://doi.org/10.1002/humu.23857>

How to cite this article: Andreoletti G, Pal LR, Moul J, Brenner SE. Reports from the fifth edition of CAGI: The Critical Assessment of Genome Interpretation. *Human Mutation*. 2019;40:1197–1201. <https://doi.org/10.1002/humu.23876>